



PERGAMON

Neuromuscular Disorders 13 (2003) 468–471



[www.elsevier.com/locate/nmd](http://www.elsevier.com/locate/nmd)

## X-inactivation patterns in carriers of X-linked myotubular myopathy

M. Kristiansen<sup>a,\*</sup>, G.P. Knudsen<sup>b</sup>, S.M. Tanner<sup>c</sup>, M. McEntagart<sup>d</sup>, H. Jungbluth<sup>e,f</sup>,  
F. Muntoni<sup>f</sup>, C. Sewry<sup>f</sup>, S. Gallati<sup>g</sup>, K.H. Ørstavik<sup>b,h</sup>, C. Wallgren-Pettersson<sup>i</sup>

<sup>a</sup>Department of Medical Genetics, Institute of Medical Genetics, University of Oslo, Oslo, Norway

<sup>b</sup>Institute Group of Clinical Medicine, University of Oslo, Oslo, Norway

<sup>c</sup>Human Cancer Genetics Program, The Ohio State University, Columbus, OH, USA

<sup>d</sup>Institute of Medical Genetics, University of Wales College of Medicine, Cardiff, Wales, UK

<sup>e</sup>Department of Paediatrics Neurology, The Newcomen Centre, Guy's Hospital, London, UK

<sup>f</sup>Dubowitz Neuromuscular Centre, Imperial College Faculty of Medicine, Hammersmith Campus, London, UK

<sup>g</sup>Department of Paediatrics, Division of Human Genetics, Inselspital, University of Berne, Berne, Switzerland

<sup>h</sup>Department of Medical Genetics, Rikshospitalet University Hospital, Oslo, Norway

<sup>i</sup>Department of Medical Genetics, University of Helsinki and Folkhälsan, Helsinki, Finland

Received 15 November 2002; received in revised form 20 February 2003; accepted 17 March 2003

### Abstract

X-linked myotubular myopathy is a rare severe muscle disorder in affected male neonates. Most female carriers are free from symptoms. Skewed X inactivation has been proposed to be responsible for the affected phenotype seen in some carriers. We have compared the X inactivation patterns in blood DNA with the clinical phenotype in carriers of X-linked myotubular myopathy. The X-inactivation analysis was performed using *Hpa*II predigestion of DNA followed by polymerase chain reaction of the highly polymorphic CAG repeat of the androgen receptor (*AR*) gene. The frequency of skewed X inactivation was similar in the X-linked myotubular myopathy carriers (22%) and in 235 controls (18%). Three overtly affected carriers had skewed X inactivation with the mutated X as the predominantly active X in at least two of them. Four females with mild symptoms had random X inactivation. The unaffected X-linked myotubular myopathy carriers had either skewed X inactivation in favour of expression from the normal X or random X-inactivation. Thus, there was a tendency for females with a more severe phenotype to have a skewed pattern of X inactivation, while females with an intermediate phenotype had a random pattern of X-inactivation.

© 2003 Published by Elsevier Science B.V.

**Keywords:** X-linked myotubular myopathy; X chromosome inactivation

### 1. Introduction

X-linked myotubular myopathy (XLMTM) is a rare congenital muscle disorder caused by mutations in the *MTM1* gene on Xq28 [1]. Most affected boys die within the first year of life because of respiratory failure [2–4]. Female carriers are usually asymptomatic or have only mild weakness [2,4–6], and inheritance has thus been regarded as X-linked recessive. However, females with severe manifestations of the disease have been reported [7–11].

One of the two X chromosomes in female mammalian cells is inactivated in early embryonic life. Females are

therefore mosaic for two different cell types, cells with the paternally inherited X chromosome as the active X, and cells with the maternally inherited X chromosome as the active X. The distribution of the two cell types normally approximates 50:50. Any marked deviation ( $\geq 80:20$ ) from this distribution is called skewed X inactivation. Older females have a higher frequency of skewed X inactivation in blood cells, probably due to a selection process [12,13].

Phenotypic variation in carriers of X-linked disorders has been attributed to a variable pattern of X-chromosome inactivation, and females affected with XLMTM have been reported showing skewed X-inactivation patterns [7,8,10,11]. In order to further investigate the relationship between X-inactivation patterns and phenotypes in carriers of XLMTM, we analysed the X-inactivation patterns in a

\* Corresponding author. Tel.: +47-230-75585; fax: +47-230-75590.  
E-mail address: marianne.kristiansen@ioks.uio.no (M. Kristiansen).

population of XLMTM carriers, non-carriers from the same families and controls.

## 2. Material and methods

### 2.1. Subjects

The series consisted of 18 XLMTM families with 43 carriers identified by mutation detection, or known by family history to be obligate carriers, and 15 non-carriers. One affected female carrier was aged 7 years. The remaining carriers were 16–71 years of age (median 37 years). Fifteen families (36 carriers) were ascertained after identification of a male patient, and three families (seven carriers) were ascertained through an affected female. Clinical information was available for 20 of the carriers (Table 1). Seven females were characterised as unaffected by history, six as unaffected by clinical examination, four as mildly affected (no history of symptomatic muscle weakness but mild weakness on examination in one or a few muscle groups, or a history of easy fatigability). Three females from three different families were overtly affected (definite subjective and objective muscle weakness for which the patient had sought medical attention) and more extensive details have been published previously [8,9,11]. The controls consisted of 235 blood donors of the same age group.

### 2.2. DNA isolation

Blood was obtained for carrier testing after informed consent and DNA was extracted from peripheral blood cells according to a standard procedure, using the automated phenol extraction method (Nucleic Acid Extractor 340A, Applied Biosystems) or using QIAGEN Maxi DNA extraction kit (QIAGEN).

### 2.3. X-inactivation analysis

X-inactivation pattern was determined by PCR of the highly polymorphic CAG repeat in the first exon of the androgen receptor (*AR*) gene [14]. Methylation of *Hpa* II sites in close proximity of this repeat correlates with X inactivation. After digestion with the methylation sensitive

enzyme *Hpa* II, a PCR product is obtained from the inactive X chromosome only. PCR products from both digested and undigested DNA were separated on an ABI 373 automated sequencer, and analysed by GeneScan software (Applied Biosystems). The X-inactivation patterns were classified as random (ratios 50:50– < 80:20) or skewed ( $\geq 80:20$ ). For XLMTM carriers with skewed X inactivation, the direction of skewing could be distinguished in carriers who had inherited the mutation from their mother. In these carriers, the maternal X is carrying the mutation while the paternal X is carrying the wild-type gene.

### 2.4. Statistical methods

The Pearson chi-squared test was used for testing categorical variables. The Fisher two-tailed exact test was used where appropriate. *P* values of less than 0.05 were taken as indicating statistical significance.

## 3. Results

The results of X-inactivation analysis in the carriers and controls are summarised in Table 1. There was no difference in the frequency of skewed X inactivation in carrier females and controls. Two overtly affected carriers were excluded from this analysis, since the families were ascertained through these affected females. Of the carriers, therefore, nine out of 41 (22%) had a skewed X inactivation as did 42 out of 235 (18%) of the controls. The non-carriers had similar X-inactivation patterns to those of the controls.

If there is a relationship between the XLMTM carrier phenotype and X-inactivation pattern, a tendency to more skewed X-inactivation patterns would be expected in carriers with more extreme phenotypes, i.e. in overtly affected carriers or in completely asymptomatic carriers. Similarly, a tendency to a more random pattern was expected in carriers with an intermediate phenotype, i.e. carriers with only slight symptoms. Such a tendency was found in our series, since none of the four mildly affected females had skewed X inactivation, whereas seven out of 16 (44%) of the overtly affected and unaffected females had skewed X-inactivation patterns. This difference, however, was not significant ( $P = 0.25$ ).

The direction of skewing was known for two of the

Table 1  
X-chromosome inactivation in XLMTM carriers, non-carriers and controls

X inactivation pattern	Carriers (%)	Carriers with clinical information				Non-carriers (%)	Controls (%)
		Unaffected (%)	Mildly affected (%)	Overtly affected (%)	Total (%)		
Random (50:50– < 80:20)	32 (74)	9 (69)	4 (100)	0 (0)	13 (65)	13 (87)	193 (82)
Skewed ( $\geq 80:20$ )	11 (26)	4 (31)	0 (0)	3 (100) [8,9,11]	7 (35)	2 (13)	42 (18)
Total	43	13	4	3	20	15	235

overtly affected females with skewed X-inactivation patterns. They had inherited the mutation from their mother as well as having the maternal, disease-carrying X chromosome, as the preferentially active X [8,9]. One of these patients had in another laboratory previously shown to have a random pattern (60:40) [9] but had a mean ratio of 83:17 in four repeated assays of the same sample in the present study. The direction of skewing was not known for the third overtly affected female [11].

One of the unaffected carriers with skewing was the mother of one of the affected females with skewing [8]. In this female, in contrast to that seen in her affected daughter, the direction of skewing was favourable, since the mutated gene was most probably on the inactive X chromosome. This was also the case for the two unaffected sisters with skewed X inactivation. They had inherited the mutation from their mother but had the paternally inherited X chromosome as the preferentially active X. The direction of skewing was not known for the other unaffected carrier with a skewed pattern.

#### 4. Discussion

Skewed X inactivation may occur due to chance (few cells at the time of X inactivation), to genetic factors influencing the X inactivation process, or to a selection process [15,16]. Carriers of several severe X-linked disorders, such as Wiskott–Aldrich syndrome, the ATR-X syndrome, some of the X-linked immunodeficiency syndromes and Barth syndrome have skewed X inactivation patterns [17–20], presumably as a result of selection against cells with the mutated gene on the active X [21]. In such disorders, female carriers will have a completely normal phenotype. Over expression of the mutated gene may cause a lethal phenotype, which may explain why affected carriers with an unfavourably skewed X inactivation have not been reported.

In many other serious X-linked disorders, including Duchenne muscular dystrophy and haemophilia, affected females have been reported with an unfavourably skewed X inactivation, with the mutated gene on the predominantly active X chromosome [22,23]. In these disorders, it is expected that no selection process takes place and that a skewed pattern occurs by chance, and is in favour either of the mutated or the normal X. Thus, under these conditions, a difference between the X inactivation pattern of carriers and controls is not expected. In the present study, no such difference was found between carriers of *MTM1* mutations and controls, which is in agreement with absence of a selection against cells expressing the X chromosome carrying the mutated *MTM1* gene.

All three overtly affected females included in this study had skewed X inactivation. Furthermore, two of them showed skewing in favour of activity of the mutated X [8,9]. The direction of skewing was not known for the third overtly affected female [11]. We also found a random X

inactivation pattern in the four carriers with mild symptoms, and skewed or random patterns in the unaffected females. For three of the four carriers with skewing and a normal phenotype, the direction of skewing was known, and was in a favourable direction.

Our findings are therefore in agreement with the proposed correlation between the X inactivation pattern and phenotype. The correlation was not significant. However, this could be due to the small number of carriers of this very rare disorder with clinical information available. However, the report of a female with XLMTM and random X inactivation both in blood and muscle DNA indicates a complex relationship between X inactivation and phenotype [10]. This has been observed in carriers of Duchenne muscular dystrophy and haemophilia also. Both skewed and random patterns were found in muscle DNA in five symptomatic carriers of Duchenne muscular dystrophy [22], and no relationship was found between the X inactivation pattern in blood and phenotype in proven carriers of haemophilia [24].

Skewed X inactivation may be due to genetic factors [15,25], and Mendelian inheritance of skewed X inactivation has been reported in families without known X-linked disorders [26,27]. Families with X-linked disorders have also been reported who have been ascertained through an affected female, where other females, both carriers and non-carriers, also had a skewed X inactivation [23,28]. This is in agreement with the findings in our study where a pair of unaffected carrier sisters both showed evidence of favourable skewed X inactivation. Furthermore, the finding of an opposite skewing pattern in an unaffected mother and her affected daughter, both of whom carry the same *MTM1* mutation, supports the existence of genetic factors influencing X inactivation patterns.

X inactivation occurs at different times in different tissues, and X inactivation patterns are therefore expected to vary between tissues [29,30]. We studied X inactivation in blood. It is possible that a more appropriate tissue, such as muscle cells, would reveal a closer correlation between X inactivation and phenotype in carriers of XLMTM.

#### Acknowledgements

This work was supported by The Research Council of Norway, and EXTRA funds from the Norwegian Foundation for Health and Rehabilitation. Meriel McEntagart and Heinz Jungbluth were supported by the Muscular Dystrophy Campaign of Great Britain. The authors are grateful to Dr David O. Robinson for providing DNA samples.

#### References

- [1] Laporte J, Hu LJ, Kretz C, et al. A gene mutated in X-linked

- myotubular myopathy defines a new putative tyrosine phosphatase family conserved in yeast. *Nat Genet* 1996;13:175–82.
- [2] Wallgren-Pettersson C, Clarke A, Samson F, et al. The myotubular myopathies: differential diagnosis of the X linked recessive, autosomal dominant, and autosomal recessive forms and present state of DNA studies. *J Med Genet* 1995;32:673–9.
- [3] Wallgren-Pettersson C. Myotubular/centronuclear myopathy. In: Emery AEH, editor. *Diagnostic criteria for neuromuscular disorders*, 2nd ed. Baarn, The Netherlands/London, UK: Royal Society of Medicine/European Neuromuscular Centre; 1997. p. 65–7.
- [4] McEntagart M, Parsons G, Buj-Bello A, et al. Genotype–phenotype correlations in X-linked myotubular myopathy. *Neuromuscul Disord* 2002;12:939–46.
- [5] Wallgren-Pettersson C. 72nd International Workshop: myotubular myopathy 1–3 October 1999, Hilversum, The Netherlands. *Neuromuscul Disord* 2000;10:525–9.
- [6] Heckmatt JZ, Sewry CA, Hodes D, Dubowitz V. Congenital centronuclear (myotubular) myopathy. A clinical, pathological and genetic study in eight children. *Brain* 1985;108:941–64.
- [7] Dahl N, Hu LJ, Chery M, et al. Myotubular myopathy in a girl with a deletion at Xq27–q28 and unbalanced X-inactivation assigns the MTM1 gene to a 600 kb region. *Am J Hum Genet* 1995;56:1108–15.
- [8] Tanner SM, Ørstavik KH, Kristiansen M, et al. Skewed X-inactivation in a manifesting carrier of X-linked myotubular myopathy and in her non-manifesting carrier mother. *Hum Genet* 1999;104:249–53.
- [9] Hammans SR, Robinson DO, Moutou C, et al. A clinical and genetic study of a manifesting heterozygote with X-linked myotubular myopathy. *Neuromuscul Disord* 2000;10:133–7.
- [10] Sutton IJ, Winer JB, Norman AN, Liechti-Gallati S, MacDonald F. Limb girdle and facial weakness in female carriers of X-linked myotubular myopathy mutations. *Neurology* 2001;57:900–2.
- [11] Jungbluth H, Sewry CA, Buj-Bello A, et al. Early and severe presentation of X-linked myotubular myopathy in a girl with skewed X-inactivation. *Neuromuscul Disord* 2003;13:55–9.
- [12] Busque L, Mio R, Mattioli J, et al. Nonrandom X-inactivation patterns in normal females: lyonization ratios vary with age. *Blood* 1996;88:59–65.
- [13] Christensen K, Kristiansen M, Hagen-Larsen H, et al. X-linked genetic factors regulate hematopoietic stem-cell kinetics in females. *Blood* 2000;95:2449–51.
- [14] Allen RC, Zoghbi HY, Moseley AB, Rosenblatt HM, Belmont JW. Methylation of HpaII and HhaI sites near the polymorphic CAG repeat in the human androgen-receptor gene correlates with X chromosome inactivation. *Am J Hum Genet* 1992;51:1229–39.
- [15] Belmont JW. Genetic control of X inactivation and processes leading to X-inactivation skewing. *Am J Hum Genet* 1996;58:1101–8.
- [16] Puck JM, Willard HF. X inactivation in females with X-linked disease. *N Engl J Med* 1998;338:325–8.
- [17] Fearon E, Kohn DB, Winkelstein JA, Vogelstein B, Blaese RM. Carrier detection in the Wiskott–Aldrich syndrome. *Blood* 1988;72:1735–9.
- [18] Gibbons RJ, Suthers GK, Wilkie OM, Buckle VJ, Higgs DR. X-linked  $\alpha$ -thalassemia/mental retardation (ATR-X) syndrome: localization to Xq12–q21.31 by X inactivation and linkage analysis. *Am J Hum Genet* 1992;51:1136–49.
- [19] Allen RC, Nachtman RG, Rosenblatt HM, Belmont JW. Application of carrier testing to genetic counseling for X-linked agammaglobulinemia. *Am J Hum Genet* 1994;54:25–35.
- [20] Ørstavik KH, Ørstavik RE, Naumova AK, et al. X chromosome inactivation in carriers of Barth syndrome. *Am J Hum Genet* 1998;63:1457–63.
- [21] Migeon BR, Haisley-Royster C. Familial skewed X inactivation and X-linked mutations: unbalanced X inactivation is a powerful means to ascertain X-linked genes that affect cell proliferation. *Am J Hum Genet* 1998;62:1555–7.
- [22] Matthews PM, Benjamin D, Van Bakel I, et al. Muscle X-inactivation patterns and dystrophin expression in Duchenne muscular dystrophy carriers. *Neuromusc Disord* 1995;5:209–20.
- [23] Ørstavik KH, Ørstavik RE, Schwartz M. Skewed X chromosome inactivation in a female with haemophilia B and in her non-carrier daughter – a genetic influence on X chromosome inactivation? *J Med Genet* 1999;36:865–6.
- [24] Ørstavik KH, Scheibel E, Ingerslev J, Schwartz M. Absence of correlation between X chromosome inactivation pattern and plasma concentration of factor VIII and factor IX in carriers of haemophilia A and B. *Thromb Haemost* 2000;83:433–7.
- [25] Plenge RM, Hendrich BD, Schwartz C, et al. A promoter mutation in the XIST gene in two unrelated families with skewed X-chromosome inactivation. *Nat Genet* 1997;17:353–6.
- [26] Hoffman EP, Pegoraro E. Skewed X inactivation can be inherited as a Mendelian trait in humans. *Am J Hum Genet Suppl* 1995;57:A49.
- [27] Naumova AK, Olien L, Bird LM, et al. Genetic mapping of X-linked loci involved in skewing of X chromosome inactivation in the human. *Eur J Hum Genet* 1998;6:552–62.
- [28] Taylor SAM, Deugau KV, Lillicrap DP. Somatic mosaicism and female-to-female transmission in a kindred with hemophilia B (factor IX deficiency). *Proc Natl Acad Sci USA* 1991;88:39–42.
- [29] Gale RE, Wheadon H, Boulos P, Linch DC. Tissue specificity of X-chromosome inactivation patterns. *Blood* 1994;83:2899–905.
- [30] Sharp A, Robinson D, Jacobs P. Age- and tissue-specific variation of X chromosome inactivation ratios in normal women. *Hum Genet* 2000;107:343–9.